

---

Product Manual

# QuickTiter™ Hepatitis C Core Antigen (HCVcAg) ELISA Kit

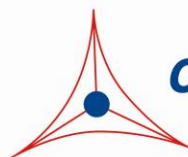
Catalog Numbers

VPK-151

96 wells

**FOR RESEARCH USE ONLY**  
**Not for use in diagnostic procedures**

---



**CELL BIOLABS, INC.**

*Creating Solutions for Life Science Research*

## **Introduction**

Hepatitis C is an infection of the liver caused by the hepatitis C virus. HCV is transmitted by exposure to infectious blood; forms of transmission include unprotected sexual activity, blood transfusion, mother-to-infant transmission, or occupational exposure to blood. The acute illness causes liver inflammation, liver fibrosis, vomiting and jaundice, while chronic HCV infection often leads to liver cirrhosis and failure.

An estimated 270-300 million people worldwide have been infected with hepatitis C. At least 75% of people infected will develop chronic hepatitis C and have it the rest of their lives.

Diagnosis of chronic hepatitis C virus (HCV) infection has long been based on HCV serology and detection of HCV antibodies. With the development of therapies for chronic HCV infection, including interferon and ribavirin, quantitative detection of HCV has been used increasingly as the most important marker for monitoring HCV titer, disease progression, and assessing antiviral treatment. Several assays for the quantitative measurement of HCV DNA have been developed, such as PCR-based nucleic acid amplification assays. However, these methods tend to be cumbersome and expensive.

Cell Biolabs' QuickTiter™ HCV Core Antigen ELISA Kit is an enzyme immunoassay developed for detection and quantitation of the HCV core protein. The kit has detection sensitivity limit of 1 ng /mL HCVcAg. Each kit provides sufficient reagents to perform up to 96 assays including standard curve and HCV samples.

## **Assay Principle**

An anti-HCVcAg monoclonal coating antibody is adsorbed onto a microtiter plate. HCV core antigen present in the sample or standard binds to the antibodies adsorbed on the plate; a FITC-conjugated mouse anti-HCVcAg antibody is added and binds to the antigen captured by the first antibody.

Following incubation and wash steps, a HRP-conjugated mouse anti-FITC antibody is added and binds to the FITC conjugated anti-HCVcAg. Unbound HRP-conjugated mouse anti-FITC antibody is removed during a wash step, and substrate solution reactive with HRP is added to the wells.

A colored product is formed in proportion to the amount of HCV core antigen present in the sample. The reaction is terminated by addition of acid and absorbance is measured at 450 nm. A standard curve is prepared from recombinant HCV core antigen and sample concentration is then determined.

## **Related Products**

1. VPK-150: QuickTiter™ HBV Core Antigen ELISA Kit
2. VPK-108-H: QuickTiter™ Lentivirus Quantitation Kit (HIV-1 p24 ELISA)
3. VPK-107: QuickTiter™ Lentivirus Titer Kit (Lentivirus-Associated HIV p24)
4. VPK-112: QuickTiter™ Lentivirus Quantitation Kit

## **Kit Components**

### **Box 1 (shipped at room temperature)**

1. Anti-HCVcAg Antibody Coated Plate (Part No. 315101): One strip well 96-well plate.
2. FITC-Conjugated Anti-HCVcAg Monoclonal Antibody (Part No. 315102): One 20  $\mu$ L vial.
3. HRP-Conjugated Anti-FITC Monoclonal Antibody (Part No. 310811): One 20  $\mu$ L vial.
4. Assay Diluent (Part No. 310804): One 50 mL bottle.
5. Triton X-100 Solution (Part No. 310805): One 15 mL bottle containing 5% Triton X-100 in TBS.
6. 10X Wash Buffer (Part No. 310806): One 100 mL bottle.
7. Substrate Solution (Part No. 310807): One 12 mL amber bottle.
8. Stop Solution (Part. No. 310808): One 12 mL bottle.

### **Box 2 (shipped on blue ice packs)**

1. Recombinant HCVcAg Standard (Part No. 315103): One 100  $\mu$ L vial of 10  $\mu$ g/mL recombinant HCV Core Antigen in 8 M Urea containing BSA.

## **Materials Not Supplied**

1. HCV Sample: purified virus or unpurified viral supernatant
2. Cell Culture Centrifuge
3. 0.45  $\mu$ m filter
4. 10  $\mu$ L to 1000  $\mu$ L adjustable single channel micropipettes with disposable tips
5. 50  $\mu$ L to 300  $\mu$ L adjustable multichannel micropipette with disposable tips
6. Multichannel micropipette reservoir
7. Microplate reader capable of reading at 450 nm (620 nm as optional reference wave length)

## **Storage**

Upon receiving, aliquot and store recombinant HCVcAg Standard at  $-20^{\circ}\text{C}$  and avoid freeze/thaw. Store all other components at  $4^{\circ}\text{C}$ .

## **Safety Considerations**

Remember that your samples contain infectious viruses before inactivation; you must follow the recommended NIH guidelines for all materials containing infectious organisms.

## **Preparation of Reagents**

- 1X Wash Buffer: Dilute the 10X Wash Buffer Concentrate to 1X with deionized water. Stir to homogeneity.

- FITC-Conjugated Anti-HCVcAg Monoclonal Antibody and HRP-Conjugated Anti-FITC Monoclonal Antibody: Immediately before use dilute the FITC-conjugated antibody 1:1000 and HRP-conjugated antibody 1:1000 with Assay Diluent. Do not store diluted solutions.

### **Preparation of Standard Curve**

1. Prepare a dilution series of Recombinant HCVcAg Standard in the concentration range of 100 ng/mL to 1 ng/mL by diluting the stock solution in Assay Diluent (Table 1).

<b>Standard Tubes</b>	<b>HCVcAg Standard (µL)</b>	<b>Assay Diluent (µL)</b>	<b>HCVcAg (ng/mL)</b>
1	10	990	100
2	500 of Tube #1	500	50
3	500 of Tube #2	500	25
4	500 of Tube #3	500	12.5
5	500 of Tube #4	500	6.25
6	500 of Tube #5	500	3.125
7	500 of Tube #6	500	1.5625
8	0	500	0

**Table 1. Preparation of HCVcAg Standard**

2. Transfer 225µL of each dilution to a microcentrifuge tube containing 25 µL of Triton X-100 Solution. Perform the assay as described in Assay Instructions.

### **HCV Sample Dilution and Inactivation**

1. (Optional) Dilute HCV sample in culture medium. Include culture medium as a negative control.
2. Transfer 225 µL of each sample to a microcentrifuge tube containing 25 µL of Triton X-100 Solution, Vortex well.
3. Incubate 30 minutes at 37°C.

*Note: For samples that contain antibodies, release HCVcAg from the virion by incubating samples at 56°C for 30 min.*

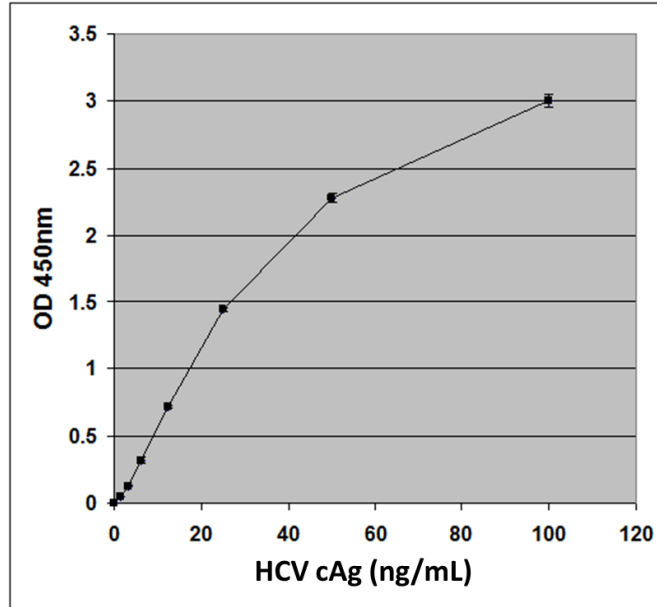
### **Assay Protocol**

1. Prepare and mix all reagents thoroughly before use.
2. Each HCV sample, HCVcAg standard, blank, and control medium should be assayed in duplicate.
3. Add 100 µL of inactivated sample or HCVcAg standard to Anti-HCVcAg Antibody Coated Plate.
4. Cover with a Plate Cover and incubate at 37°C for 2 hours.

5. Remove plate cover and empty wells. Wash microwell strips 5 times with 250  $\mu$ L 1X Wash Buffer per well with thorough aspiration between each wash. After the last wash, empty wells and tap microwell strips on absorbent pad or paper towel to remove excess 1X Wash Buffer.
6. Add 100  $\mu$ L of the diluted FITC-Conjugated Anti-HCVcAg Monoclonal Antibody to each well.
7. Cover with a plate cover and incubate at room temperature for 1 hour on an orbital shaker.
8. Remove plate cover and empty wells. Wash the strip wells 5 times according to step 5 above.
9. Add 100  $\mu$ L of the diluted HRP-Conjugated Anti-FITC Monoclonal Antibody to all wells.
10. Cover with a plate cover and incubate at room temperature for 1 hour on an orbital shaker.
11. Remove plate cover and empty wells. Wash microwell strips 5 times according to step 5 above. Proceed immediately to the next step.
12. Warm Substrate Solution to room temperature. Add 100  $\mu$ L of Substrate Solution to each well, including the blank wells. Incubate at room temperature on an orbital shaker. Actual incubation time may vary from 5-20 minutes.  
  
*Note: Watch plate carefully; if color changes rapidly, the reaction may need to be stopped sooner to prevent saturation.*
13. Stop the enzyme reaction by adding 100  $\mu$ L of Stop Solution into each well, including the blank wells. Results should be read immediately (color will fade over time).
14. Read absorbance of each microwell on a spectrophotometer using 450 nm as the primary wave length.

### **Example of Results**

The following figures demonstrate typical HCVcAg ELISA results. One should use the data below for reference only. This data should not be used to interpret actual results.



**Figure 1: HCV Core Antigen ELISA Standard Curve**

## **References**

1. Alter, H. J., R. H. Purcell, J. W., Shih, J. C. Melpolder, M. Houghton, Q.-L. Choo, and G. Kuo (1989) *N. Engl. J. Med.* 321:1494-1500.
2. Choo, Q.-L., G. Kuo, A. J. Weiner, L. R. Overby, D. W. Bradley, and M. Houghton (1989) *Science* 244:359-362.
3. Kuo, G., Q.-L. Choo, H. J. Alter, G. I. Gitnick, A. G. Redeker, R. H. Purcell, T. Miyamura, J. L. Dienstag, M. J. Alter, C. E. Stevens, G. E. Tegtmeier, F. Bonino, M. Colombo, W. S. Lee, C. Kuo, K. Berger, J. R. Shuster, L. R. Overby, D. W. Bradley, and M. Houghton (1989) *Science* 244:362-364.
4. Alias, M. B., K. Patel, H. Dahari, S. Beaucourt, P. Larderie, L. Blatt, C. Hezode, G. Picchio, D. Dhumeaux, A. U. Neumann, J. G. McHutchison, and J. M. Pawlotsky (2002) *Hepatology* 36:211-218.
5. Aoyagi, K., C. Ohue, K. Iida, T. Kimura, E. Tanaka, K. Kiyosawa, and S. Yagi (1999) *J. Clin. Microbiol.* 37:1802-1808.
6. Kurtz, J. B., E. Boxall, N. Qusir, J. Shirley, D. Coleman, and C. Chandler (2001) *J. Virol. Methods* 96:127-132.

## **Recent Product Citations**

1. Rani, S.V. et al. (2023). Detection of Hepatitis C Virus Infection and its Genotypic Characterisation among Hemodialysis Patients by HCV Core Antigen Elisa and RTPCR- A Hospital- Based Prospective Study in a Tertiary Care Hospital in South India. *Int J Acad Med Pharm.* 5(4): 628-632. doi: 10.47009/jamp.2023.5.4.125.
2. Kong, L. et al. (2021). The synthetic opioid fentanyl enhances viral replication in vitro. *PLoS One.* 16(4):e0249581. doi: 10.1371/journal.pone.0249581.
3. Hammerstad, S.S. et al. (2019). Hepatitis C virus infection of human thyrocytes: metabolic, hormonal, and immunological implications. *J Clin Endocrinol Metab.* pii: dgz241. doi: 10.1210/clinem/dgz241.

4. Blackard, J.T. et al. (2019). CCR5 receptor antagonism inhibits hepatitis C virus (HCV) replication in vitro. *PLoS One*. **14**(10):e0224523. doi: 10.1371/journal.pone.0224523.
5. Plissonnier, M.L. et al. (2019). LARP1 binding to hepatitis C virus particles is correlated with intracellular retention of viral infectivity. *Virus Res*. doi: 10.1016/j.virusres.2019.197679.
6. Kumar, S. et al. (2019). MARCH8 Ubiquitinates the Hepatitis C Virus Nonstructural 2 Protein and Mediates Viral Envelopment. *Cell Rep*. **26**(7):1800-1814.e5. doi: 10.1016/j.celrep.2019.01.075.
7. Hassan, R. et al. (2018). Role of Hepatitis C Virus Core Antigen Assay in Blood Donors Screening at Zagazig University Hospitals. *Afro-Egyptian Journal of Infectious and Endemic Diseases*. **8**(1):48-54. doi: 10.21608/aeji.2018.8735.
8. Xiao, F. et al. (2018). Interactions between the Hepatitis C Virus Nonstructural 2 Protein and Host Adaptor Proteins 1 and 4 Orchestrate Virus Release. *MBio*. **9**(2). pii: e02233-17. doi: 10.1128/mBio.02233-17.
9. Blackard, J.T. et al. (2017). A preliminary analysis of hepatitis C virus in pancreatic islet cells. *Virol J*. **14**(1):237. doi: 10.1186/s12985-017-0905-3.
10. Dvir, R. et al. (2017). Autoimmune hepatitis and occult HCV infection: A prospective single-centre clinical study. *Autoimmun Rev*. **16**(3):323-325. doi: 10.1016/j.autrev.2017.01.015.
11. Owolabi, O. B. et al. (2015). Hepatitis C virus seroprevalence, antigenemia and associated risk factors among pregnant women in Nigeria. *Ethiop Med J*. **53**.
12. Liu, D. et al. (2015). Downregulation of miRNA-30c and miR-203a is associated with Hepatitis C virus core protein-induced epithelial-mesenchymal transition in normal hepatocytes and hepatocellular carcinoma cells. *Biochem Biophys Res Commun*. doi: 10.1016/j.bbrc.2015.07.107.
13. Jittavisutthikul, S. et al. (2015). Humanized-VHH transbodies that inhibit HCV protease and replication. *Viruses*. **7**:2030-2056.
14. Amet, T. et al. (2014). BST-2 expression in human hepatocytes is inducible by all three types of interferons and restricts production of hepatitis C virus. *Curr Mol Med*. **14**:349-360.
15. Kong, L. et al. (2014). HIV infection of hepatocytes results in a modest increase in hepatitis C virus expression in vitro. *PLoS One*. **9**:e83728.
16. Carpentier, A. et al. (2014). Engrafted human stem cell-derived hepatocytes establish an infectious HCV murine model. *J Clin Invest*. **124**:4953-4964.
17. Deng, A. et al. (2014). Human immunodeficiency virus type 1 Vpr increases hepatitis C virus RNA replication in cell culture. *Virus Res*. **184**:93-102.

## **Warranty**

These products are warranted to perform as described in their labeling and in Cell Biolabs literature when used in accordance with their instructions. THERE ARE NO WARRANTIES THAT EXTEND BEYOND THIS EXPRESSED WARRANTY AND CELL BIOLABS DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR WARRANTY OF FITNESS FOR PARTICULAR PURPOSE. CELL BIOLABS's sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of CELL BIOLABS, to repair or replace the products. In no event shall CELL BIOLABS be liable for any proximate, incidental or consequential damages in connection with the products.

## **Contact Information**

Cell Biolabs, Inc.  
7758 Arjons Drive  
San Diego, CA 92126  
Worldwide: +1 858-271-6500  
USA Toll-Free: 1-888-CBL-0505  
E-mail: [tech@cellbiolabs.com](mailto:tech@cellbiolabs.com)  
[www.cellbiolabs.com](http://www.cellbiolabs.com)

©2010-2024: Cell Biolabs, Inc. - All rights reserved. No part of these works may be reproduced in any form without permissions in writing.