
Product Manual

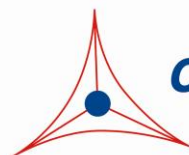
2-Phosphoglycerate Assay Kit

Catalog Number

MET-5167

100 assays

FOR RESEARCH USE ONLY
Not for use in diagnostic procedures



CELL BIOLABS, INC.

Creating Solutions for Life Science Research

Introduction

2-Phosphoglycerate (2-PG) is a glyceric acid which serves as an intermediate in the metabolic process known as glycolysis. The enzyme phosphoglycerate mutase converts 3-phosphoglycerate to 2-PG, while the enzyme enolase catalyzes 2-PG into phosphoenolpyruvate (PEP), the second to the last step in the conversion of glucose to pyruvate. In bacteria, 2-PG molecules covalently attach to enolase which causes the enzyme to be inactive. Overexpression of phosphoglycerate mutase is linked with tumor growth, survival, and invasion in many types of human cancers.

Cell Biolabs' 2-Phosphoglycerate Assay Kit is a simple fluorometric assay that measures the amount of 2-PG present in biological samples in a 96-well microtiter plate format. Each kit provides sufficient reagents to perform up to 100 assays*, including blanks, 2-PG standards, and unknown samples. Sample 2-PG concentrations are determined by comparison with a known 2-PG standard. The kit has a detection sensitivity limit of 3.1 μM 2-PG.

****Note: Each sample replicate requires 2 assays, one treated with Enolase (+E) and one without (-E). 2-PG is calculated from the difference in RFU readings from the 2 wells.***

Assay Principle

Cell Biolabs' 2-Phosphoglycerate Assay Kit measures total 2-PG within biological samples. 2-PG is converted by Enolase to phosphoenolpyruvate (PEP). PEP in the presence of ADP is converted by Pyruvate Kinase to pyruvate + adenosine triphosphate (ATP). Pyruvate is converted by pyruvate oxidase in the presence of phosphate and oxygen into acetyl phosphate, carbon dioxide, and hydrogen peroxide. The resulting hydrogen peroxide is then detected with a highly specific fluorometric probe. Horseradish peroxidase catalyzes the reaction between the probe and hydrogen peroxide, which bind in a 1:1 ratio. Samples are compared to a known concentration of 2-PG standard within the 96-well microtiter plate format. Samples and standards are incubated for 60 minutes and then read with a standard 96-well fluorometric plate reader (Figure 1).

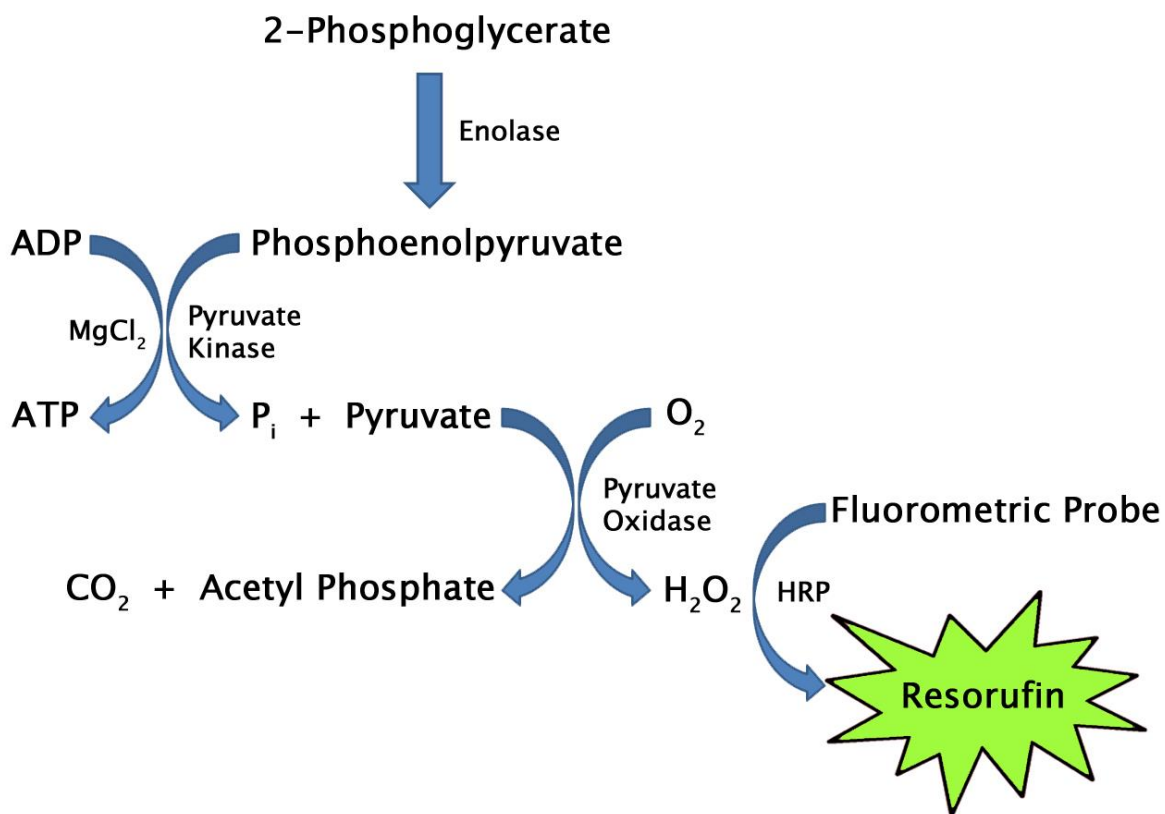


Figure 1. 2-PG Assay Principle

Related Products

1. MET-5162: Phosphoenolpyruvate Assay Kit
2. MET-5159: Pyrophosphate Assay Kit
3. MET-5029: Pyruvate Assay Kit (Fluorometric)
4. MET-5158: Methionine Assay Kit
5. MET-5152: s-Adenosylmethionine ELISA Kit
6. MET-5151: s-Adenosylhomocysteine ELISA Kit
7. MET-5001: Lactose Assay Kit
8. MET-5013: Lactate Assay Kit (Fluorometric)
9. MET-5023: Glycogen Assay Kit (Fluorometric)
10. STA-399: Free Glycerol Assay Kit (Fluorometric)
11. STA-674: Glutamate Assay Kit
12. STA-675: Hydroxyproline Assay Kit

Kit Components

Box 1 (shipped on blue ice packs)

1. 2-PG Standard (Part No. 51671C): One 50 μ L tube of 2-Phosphoglycerate at 20 mM.
2. Enolase (Part No. 51672D): One 50 μ L tube of 20 U/mL.
3. ADP (Part No. 51624C): One 250 μ L tube of adenosine diphosphate (ADP) at 200 mM.
4. Fluorometric Probe (Part No. 50231C): One 50 μ L tube in DMSO.
5. HRP (Part No. 234402-T): One 10 μ L tube of a 100 U/mL solution in glycerol.
6. FAD (Part No. 50293C): One 50 μ L tube of 2 mM Flavin Adenine Dinucleotide (FAD).
7. TPP (Part No. 50294C): One 50 μ L tube of 2 mM Thiamine Pyrophosphate (TPP).
8. Pyruvate Oxidase (Part No. 50295C): One 300 μ L tube.
9. Pyruvate Kinase (Part No. 51625D): One 200 μ L tube.

Box 2 (shipped on blue ice packs)

1. 10X Assay Buffer (Part No. 51622A): One 25 mL bottle.
2. MgCl₂ (Part No. 51623A): One 200 μ L tube of magnesium chloride at 1 M.

Materials Not Supplied

1. 10 kDa molecular weight cutoff (MWCO) centrifuge spin filter (e.g., Amicon Ultra 0.5mL)
2. 10 μ L to 1000 μ L adjustable single channel micropipettes with disposable tips
3. 50 μ L to 300 μ L adjustable multichannel micropipette with disposable tips
4. Standard 96-well fluorescence black microtiter plate and/or black cell culture microplate
5. Multichannel micropipette reservoir
6. Fluorescence microplate reader capable of reading excitation in the 530-570 nm range and emission in the 590-600 nm range.

Storage

Upon receipt, store the 10X Assay Buffer and the MgCl₂ at room temperature. Store the Pyruvate Kinase and Enolase at -80°C. Store all other components at -20°C. The Fluorometric Probe is light sensitive and must be stored accordingly. Avoid multiple freeze/thaw cycles.

Preparation of Reagents

- 1X Assay Buffer: Dilute the 10X Assay Buffer 1:10 with deionized water for a 1X solution. Stir or vortex to homogeneity. Store at 4°C.
- Reaction Mix and Negative Control Mix: Prepare two separate mixtures according to the table below. The Enolase is omitted from the Negative Control Mix.

Component	Reaction Mix (20 assays)	Negative Control Mix (20 assays)
Enolase	10 μ L	-----
Pyruvate Kinase	40 μ L	40 μ L
MgCl ₂	40 μ L	40 μ L
ADP	50 μ L	50 μ L
HRP	2 μ L	2 μ L
Pyruvate Oxidase	60 μ L	60 μ L
FAD	10 μ L	10 μ L
TPP	10 μ L	10 μ L
Fluorometric Probe	10 μ L	10 μ L
1X Assay Buffer	768 μ L	778 μ L
Total	1000 μL	1000 μL

Note: Prepare only enough for immediate use and scale proportionally as needed.

Preparation of Samples

- Cell culture supernatants: Cell culture media formulated with pyruvate should be avoided. To remove insoluble particles, centrifuge at 10,000 x g for 5 min. Collect the supernatant and filter the solution with a 10 kDa spin filter to deproteinate the sample. Collect flow through. The flow through may be assayed directly or diluted as necessary into PBS.

Note: Maintain pH between 7 and 8 for optimal working conditions as the Fluorometric Probe is unstable at high pH (>8.5).

- Tissue lysates: Sonicate or homogenize 100-500 milligrams of tissue sample in PBS and centrifuge at 10,000 x g for 10 minutes at 4°C. Collect the supernatant and filter the solution with a 10 kDa spin filter to deproteinate the sample. Collect flow through. The flow through may be assayed directly or diluted as necessary into PBS.
- Cell lysates: Resuspend cells at 1-2 x 10⁶ cells/mL in PBS. Homogenize or sonicate the cells on ice. Centrifuge to remove debris at 10,000 x g for 10 minutes at 4°C. Collect the supernatant and filter the solution with a 10 kDa spin filter to deproteinate the sample. Collect flow through. The flow through may be assayed undiluted or diluted as necessary into PBS.
- Serum, plasma, saliva, or urine: To remove insoluble particles, centrifuge at 10,000 x g for 10 minutes. Collect the supernatant and filter the solution with a 10kDa spin filter to deproteinate the sample. Collect flow through. The flow through may be assayed directly or diluted as necessary into PBS.

Notes:

- All samples should be assayed immediately or stored at -80°C for up to 1-2 months. Run proper controls as necessary. Optimal experimental conditions for samples must be determined by the investigator. Always run a standard curve with samples.
- Samples with NADH concentrations above 10 µM and glutathione concentrations above 50 µM will oxidize the Fluorometric Probe and could result in erroneous readings. To minimize this interference, it is recommended that superoxide dismutase (SOD) be added to the reaction at a final concentration of 40 U/mL (Votyakova and Reynolds, Ref. 2).
- Avoid samples containing DTT or β-mercaptoethanol since the Fluorometric Probe is not stable in the presence of thiols (above 10 µM).

Preparation of Standard Curve

Prepare fresh 2-PG standards by diluting into 1X Assay Buffer according to Table 1 below.

Standard Tubes	20 mM 2-PG Solution (µL)	1X Assay Buffer (µL)	2-PG (µM)
1	5	495	200
2	250 of Tube #1	250	100
3	250 of Tube #2	250	50
4	250 of Tube #3	250	25
5	250 of Tube #4	250	12.5
6	250 of Tube #5	250	6.25
7	250 of Tube #5	250	3.13
8	0	250	0

Table 1. Preparation of 2-PG Standards.

Assay Protocol

1. Prepare and mix all reagents thoroughly before use. Each sample, including unknowns and standards, should be assayed in duplicate or triplicate.
Note: Each unknown sample replicate requires two paired wells, one to be treated with Enolase (+E) and one without the enzyme (-E) to measure endogenous background.
2. Add 50 µL of each 2-PG standard or unknown sample into wells of a 96-well microtiter plate.
3. Add 50 µL of Reaction Mix to the standards and to one half of the paired sample wells, and mix the well contents thoroughly.
4. Add 50 µL of Negative Control Mix to the other half of the paired sample wells.
5. Mix the well contents thoroughly and incubate for 60 minutes at 37°C protected from light.
Note: This assay is continuous (not terminated) and therefore may be measured at multiple time points to follow the reaction kinetics.
6. Read the plate with a fluorescence microplate reader equipped for excitation in the 530-570 nm range and for emission in the 590-600 nm range.

Calculation of Results

1. Determine the average Relative Fluorescence Unit (RFU) values for each sample, control, and standard.
2. Subtract the average zero standard value from itself and all standard values.
3. Graph the standard curve (see Figure 2).
4. Subtract the sample well values without Enolase (-E) from the sample well values containing Enolase (+E) to obtain the difference. The fluorescence difference is due to the Enolase activity.

$$\text{Net RFU} = (\text{RFU}_{+E}) - (\text{RFU}_{-E})$$

5. Compare the net RFU of each sample to the standard curve to determine and extrapolate the quantity of 2-PG present in the sample. Only use values within the range of the standard curve.

Example of Results

The following figures demonstrate typical 2-Phosphoglycerate Assay Kit results. One should use the data below for reference only. This data should not be used to interpret or calculate actual sample results.

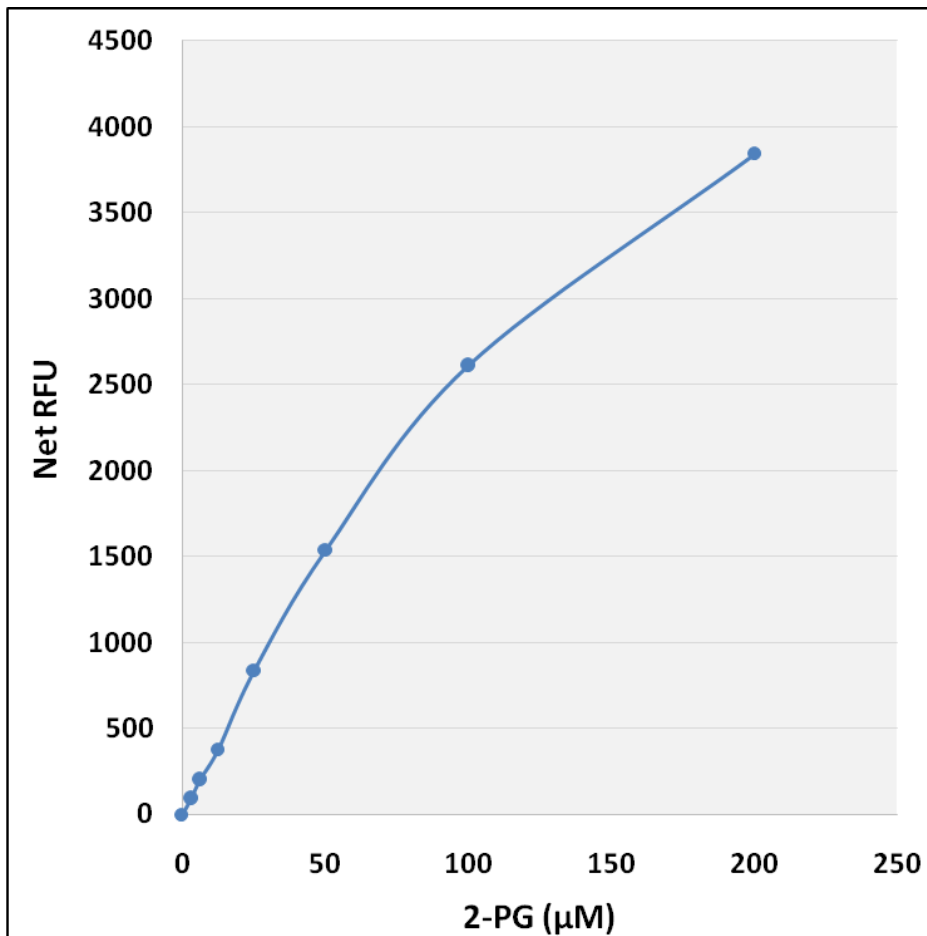


Figure 2: 2-PG Standard Curve.

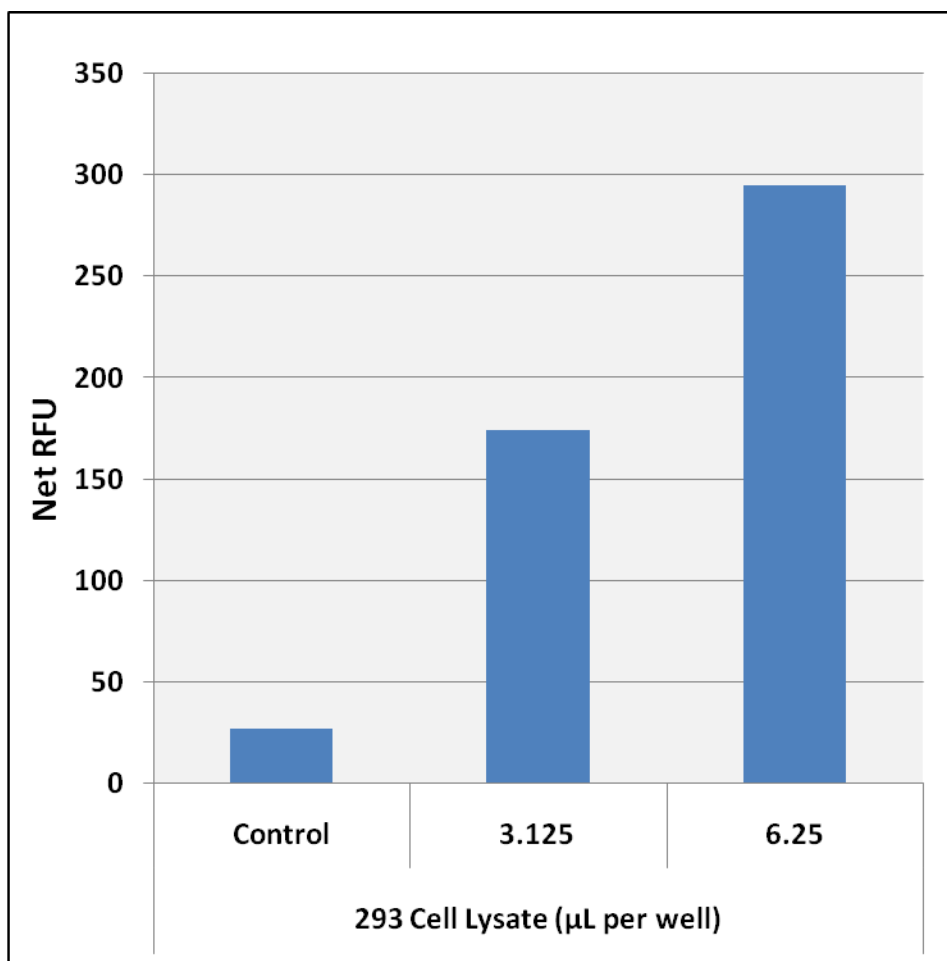


Figure 3: 2-PG detection in 293 Cells using the 2-Phosphoclycerate Assay Kit (Fluorometric). Cells were homogenized and deproteinated according to the preparation of samples section above.

References

1. Boël G Pichereau V Mijakovic I Mazé A Poncet S Gillet S Giard J-C Hartke A Auffray Y and Deutscher J (2004) *J. Mol. Bio.* **337**:485-496
2. Votyakova TV, and Reynolds IJ (2001) *J. Neurochem.* **79**:266-277.
3. Sharif F, Rasul A, Ashraf A, Hussain G, Younis T, Sarfraz I, Chaudhry MA, Bukhari SA, Ji XY, Selamoglu Z, and Ali M (2019) *Int. U. Biochem. Mol. Biol.* **71**:1418-1427.

Warranty

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